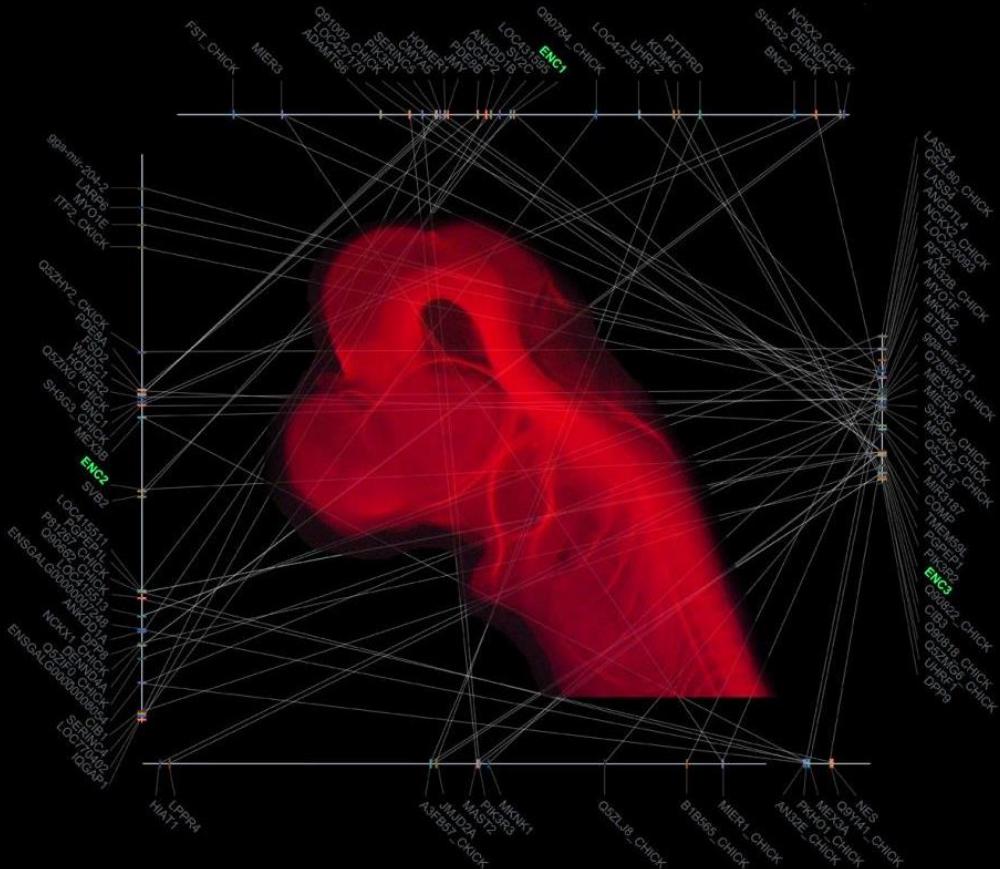




# Assessing vertebrate homolog space ... from Evo-Devo viewpoint



Catshark embryo with neuronal staining  
on ENC1-related tetra-synteny blocks

Shigehiro Kuraku

Unit Leader  
Phyloinformatics Unit  
RIKEN CLST (Twitter: @clst\_gras)

Adjunct Associate Prof.  
Kwansei Gakuin University

Adjunct Associate Prof.  
Kobe University

# NGS & Phyloinfo in Kobe

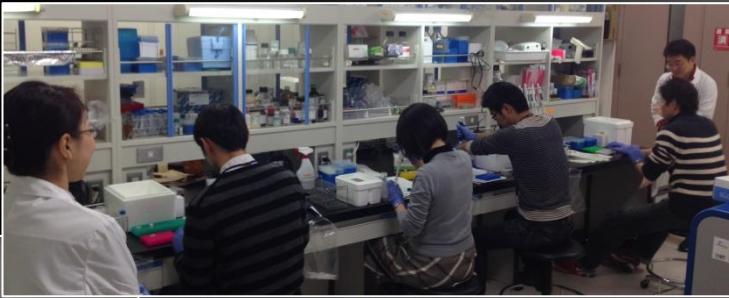
## More than just a facility

- Strength in ChIP-seq/ChIP-exo & non-model vertebrates
- Experienced with limited samples
- Cost-saving protocols (e.g. iMate Protocol, Tatsumi *et al.*, 2015)
- Flexible project consultation for biologists

Illumina HiSeq 1500



Orbitrap Velos Pro  
for LC-MS/MS



NGS Library Prep Hands-on tutorial  
January 20/21, 2015



Data analysis tutorial 'Sequence Informatics Afternoon'  
April 23, 2014

- Hands-on tutorials (ChIP, data analysis, ...)
- Organizing meetings etc.
- Facility management consultation

# Biological scope

Taxon: Vertebrata

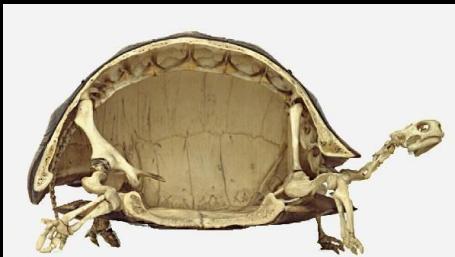
Genes: regulatory genes (TFs, signaling ligands)

What in genome allowed characters unique to vertebrates?

e.g., organized brain, visceral organs, fins/limbs, ....

What molecular change triggered morphological evolution  
in vertebrates?

Turtle shell

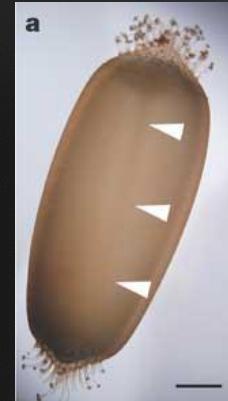


Lamprey head



<http://gallery.nanfa.org/>

Hagfish embryo



Ota, Kuraku, & Kuratani  
2007. *Nature*

<https://whyevolutionisttrue.wordpress.com>

# Orthology: thoughts and concerns

Experience through collaborations with non-experts

‘Conservation of toolkit’ overtrusted  
*(Sean Carroll, 2005. etc.)*

Is ‘Ortholog’ a jargon?

Tree-based methods underappreciated (cf. synteny)

Why?

Inaccessibility to experts’ instructions

No established standard method for tree inference

Sequence information scattered in many DBs

Tolerant peer review system for phylogenetics

# Orthology: expectations!!



Phylogeny to phenotypic evolution

:  
:  
:

Inter-family co-evolution

e.g., massive loss of embryonic axis formation genes in mammals

*Kuraku & Kuratani., 2011. GBE*

Application of phylogeny-based approaches

Phylogeny-based genome scaffolding

e.g., ESPRIT *Dessimoz et al., 2011. Brief. Bioinfo.*

# Agenda

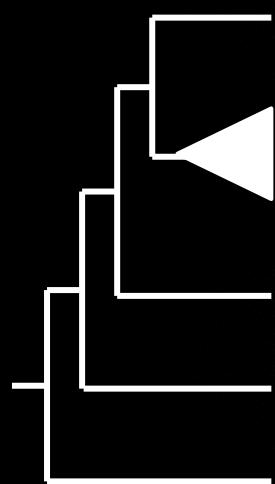
- 1) Revealing non-conservation of developmental ‘toolkits’  
‘Cryptic pan-vertebrate genes’  
=> markers for quality assessment of orthology predictions?
- 2) ‘Two-round’ whole genome duplications  
Warning in using lamprey genomes  
‘lamprey dialect’ misleading phylogeny inference?  
access to less incomplete predicted gene set

# Cryptic pan-vertebrate genes analyzed by my team

**Hox14** – Feiner et al, 2011. J Exp Zool B & Kuraku et al, 2008. PNAS

**Bmp16** – Feiner et al, 2008. BMC Evol Biol

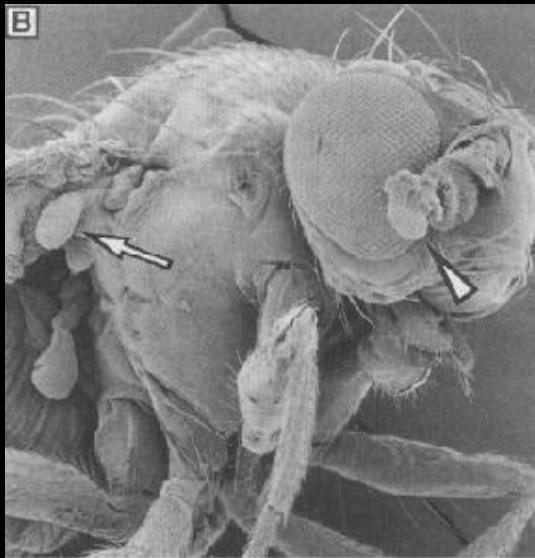
**Pax4** & **Pax10** – Feiner et al, 2014. GBE & Manousaki et al, 2011. Evol Dev



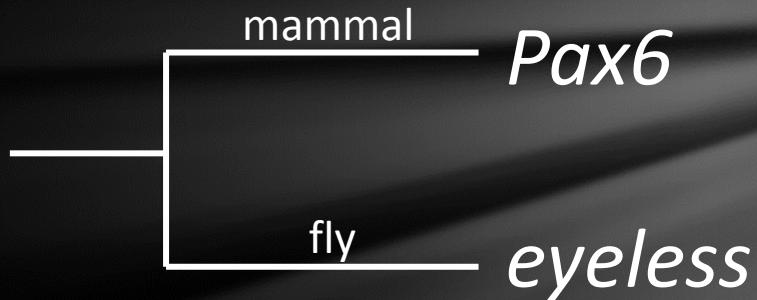
Taxon	<i>Hox14</i>	<i>Bmp16</i>	<i>Pax4</i>	<i>Pax10</i>
Mammals	–	–	+	–
Birds	–	–	–	–
Reptiles	–	+	+	+
Amphibians	–	–	–	+
Actinopterygian fish	–	+	+	+
Cartilaginous fish	+	+	?	+

# *Pax6*-*eyeless* orthology

Ectopic eyes in fly



Halder *et al.*, 1995. *Science*



# Limitation of conventional methods

0.2 substitutions / site

PhyML

99 aa

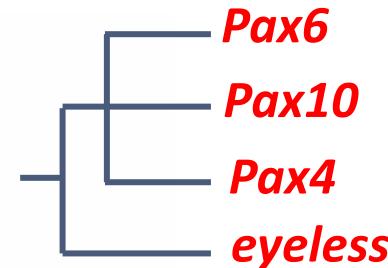
JTT+G4 ( $\alpha=0.57$ )

ML BP / Bayesian PP  
on heuristic ML tree

76/0.96  
Fugu pax6a  
medaka pax6a  
stickleback pax6a  
zebrafish pax6a  
zebrafish pax6b  
Fugu pax6b  
stickleback pax6b  
medaka pax6b  
spotted gar  
elephant shark  
human  
*X. laevis*  
chicken  
mouse  
cow  
*Scyliorhinus canicula*  
opossum

Teleost  
*pax6*  
Jawed  
vertebrate  
*Pax6*

**Pax6**



Vertebrate  
*Pax10*

**Pax10**

Invertebrate Pax6/eyeless

**Invertebrate  
Pax6/eyeless**

stickleback  
Fugu  
medaka  
zebrafish  
Alligator  
platypus

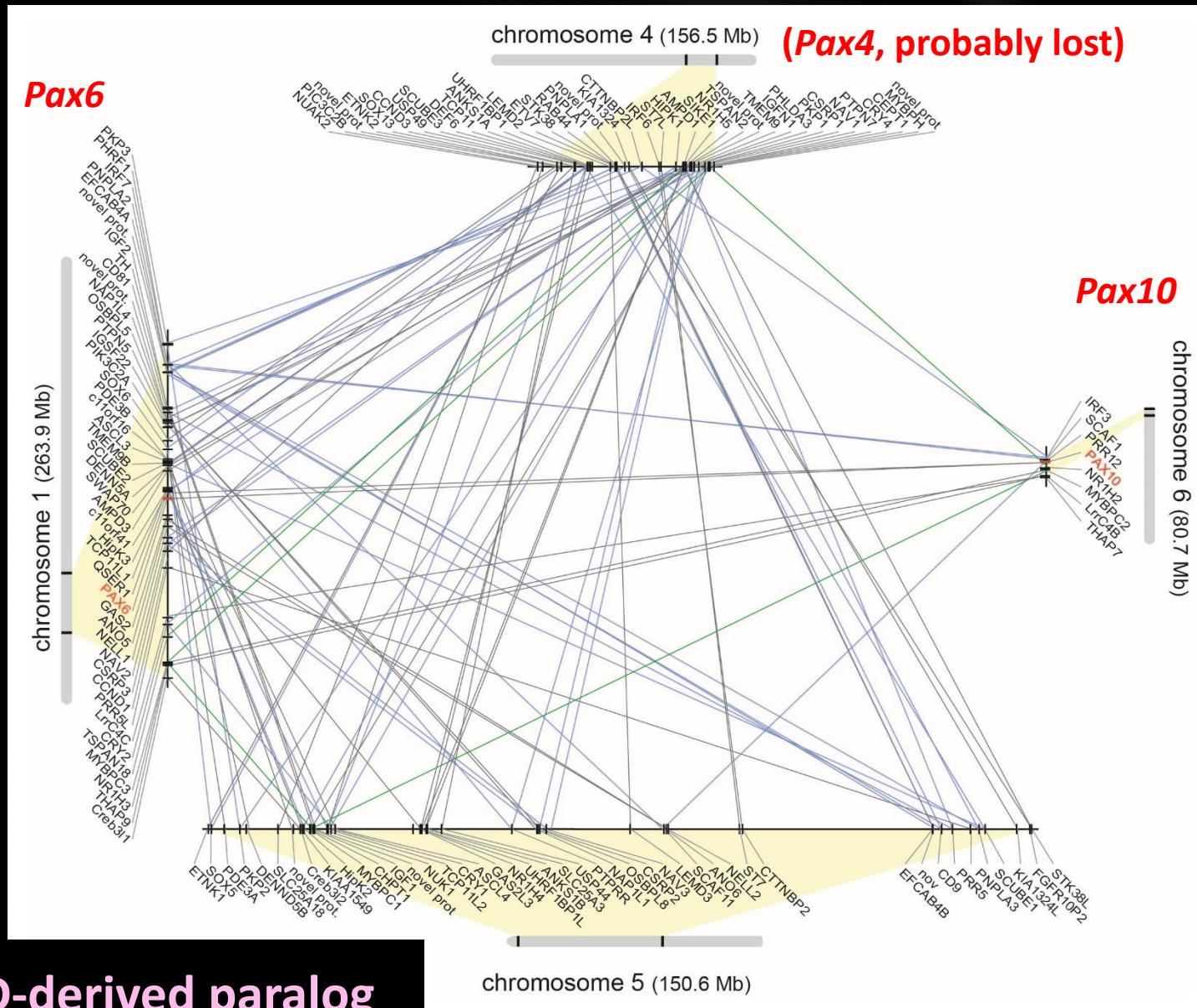
**Pax4**

99/1.00  
*Nematostella vectensis*  
fly paired  
*Ciona intestinalis Pax3/7*  
74/-  
human Pax3  
human Pax7

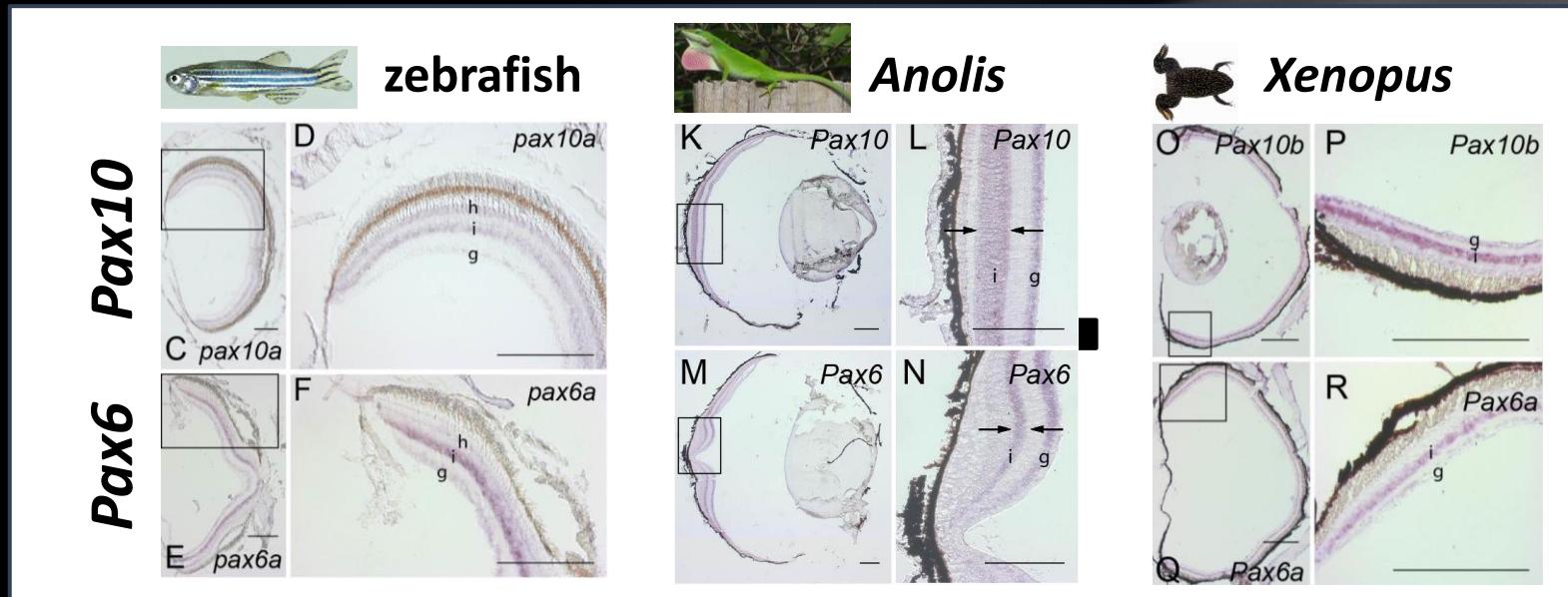
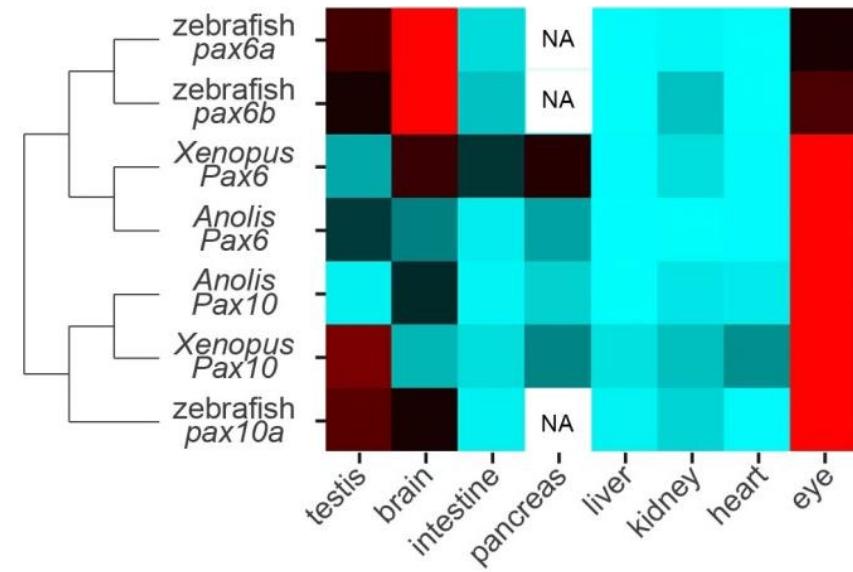
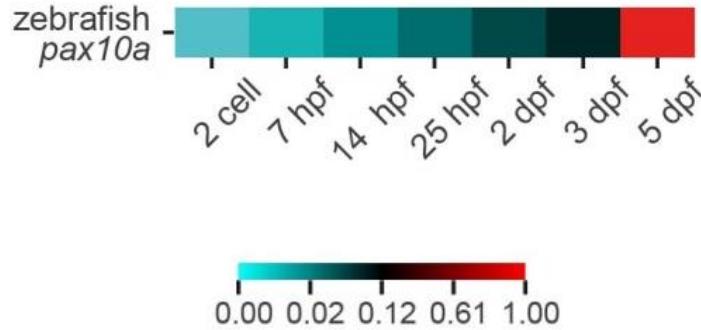
Invertebrate Pax6/eyeless

**Invertebrate  
Pax6/eyeless**

# Conserved synteny in anole lizard genome



# *Pax10*, not a useless gene

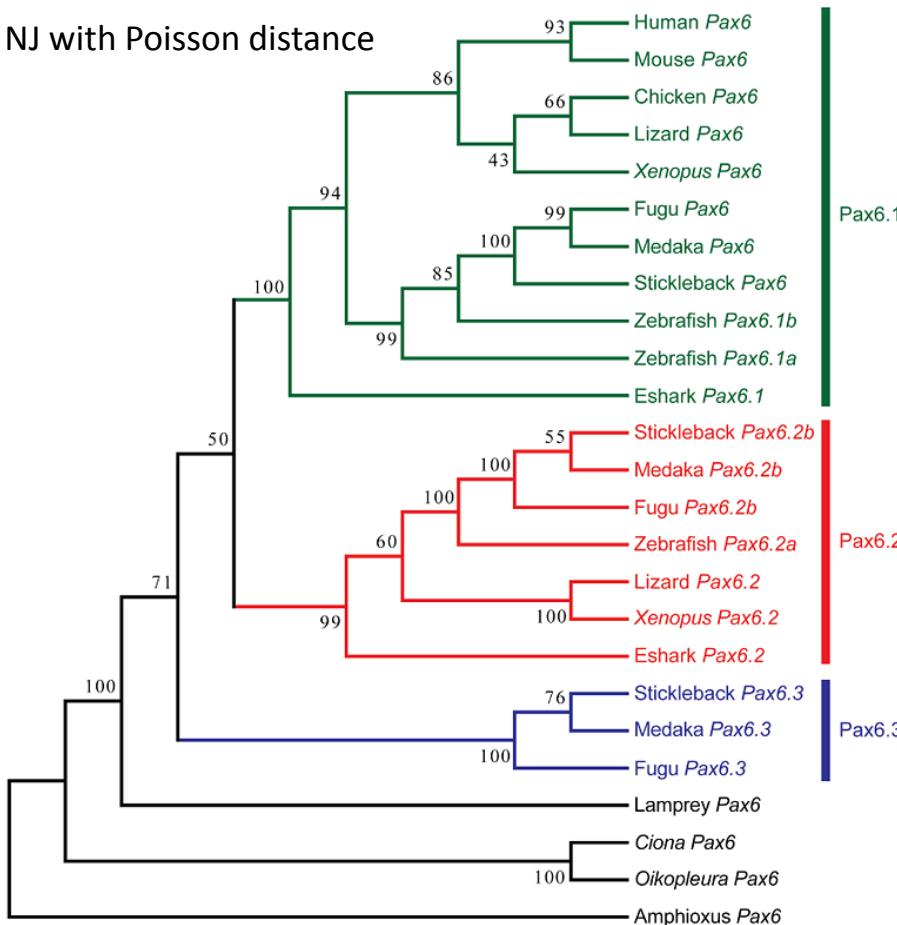


## Sequencing of *Pax6* Loci from the Elephant Shark Reveals a Family of *Pax6* Genes in Vertebrate Genomes, Forged by Ancient Duplications and Divergences

Vydianathan Ravi<sup>1\*</sup>, Shipra Bhatia<sup>2\*</sup>, Philippe Gautier<sup>2</sup>, Felix Loosli<sup>3</sup>, Boon-Hui Tay<sup>1</sup>, Alice Tay<sup>1</sup>, Emma Murdoch<sup>2</sup>, Pedro Coutinho<sup>2</sup>, Veronica van Heyningen<sup>2</sup>, Sydney Brenner<sup>1</sup>, Byrappa Venkatesh<sup>1\*</sup>, Dirk A. Kleinjan<sup>2\*</sup>

**1** Institute of Molecular and Cell Biology, Agency for Science Technology and Research (A\*STAR), Biopolis, Singapore, Singapore, **2** MRC Human Genetics Unit, MRC Institute of Genetics and Molecular Medicine, University of Edinburgh, Edinburgh, United Kingdom, **3** Institute of Toxicology and Genetics, Karlsruhe Institute of Technology, Eggenstein-Leopoldshafen, Germany

### NJ with Poisson distance



## Evolution of the Vertebrate Pax4/6 Class of Genes with Focus on Its Novel Member, the *Pax10* Gene

Nathalie Feiner<sup>1,2,3</sup>, Axel Meyer<sup>1,2</sup>, and Shigehiro Kuraku<sup>1,2,4,\*</sup>

<sup>1</sup>Department of Biology, University of Konstanz, Germany

<sup>2</sup>International Max-Planck Research School (IMPRS) for Organismal Biology, University of Konstanz, Germany

<sup>3</sup>Present address: Department of Zoology, University of Oxford, United Kingdom

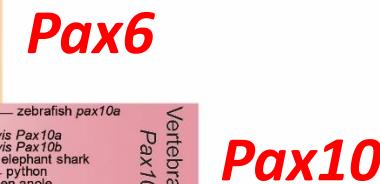
<sup>4</sup>Present address: Genome Resource and Analysis Unit, RIKEN Center for Developmental Biology, Chuo-ku, Kobe, Hyogo, Japan

\*Corresponding author: E-mail: shigehiro-kuraku@cdb.riken.jp.

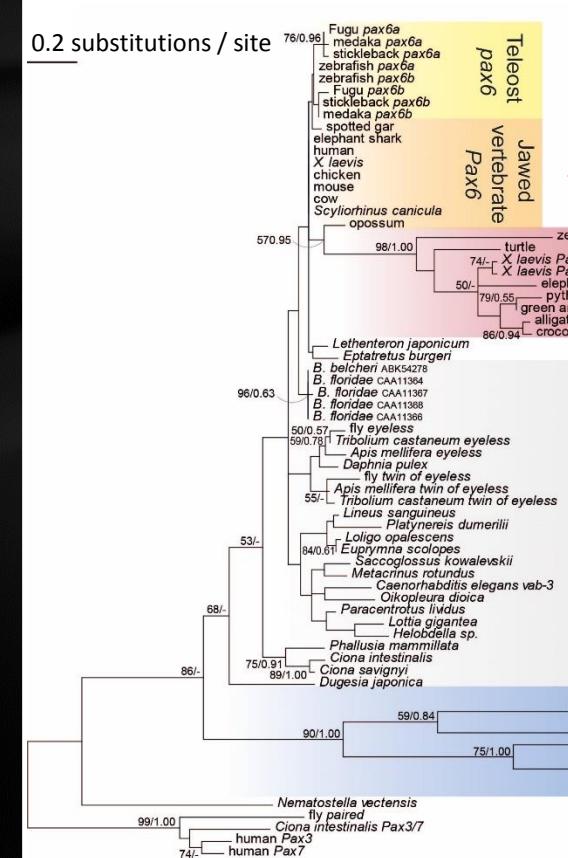
Accepted: June 14, 2014

Data deposition: This project has been deposited at EMBL under the accessions HF567444–HF567455.

ML BP / Bayesian PP  
on heuristic ML tree



Invertebrate  
Pax6/eyeless



Pax4

## Sequencing of *Pax6* Loci from the Elephant Shark Reveals a Family of *Pax6* Genes in Vertebrate Genomes, Forged by Ancient Duplications and Divergences

Vydianathan Ravi<sup>1\*</sup>, Shipra Bhatia<sup>2\*</sup>, Philippe Gautier<sup>2</sup>, Felix Loosli<sup>3</sup>, Boon-Hui Tay<sup>1</sup>, Alice Tay<sup>1</sup>, Emma Murdoch<sup>2</sup>, Pedro Coutinho<sup>2</sup>, Veronica van Heyningen<sup>2</sup>, Sydney Brenner<sup>1</sup>, Byrapa Venkatesh<sup>1\*</sup>, Dirk A. Kleinjan<sup>2\*</sup>

**1** Institute of Molecular and Cell Biology, Agency for Science Technology and Research (A\*STAR), Biopolis, Singapore, Singapore, **2** MRC Human Genetics Unit, MRC Institute of Genetics and Molecular Medicine, University of Edinburgh, Edinburgh, United Kingdom, **3** Institute of Toxicology and Genetics, Karlsruhe Institute of Technology, Eggenstein-Leopoldshafen, Germany

## Evolution of the Vertebrate Pax4/6 Class of Genes with Focus on Its Novel Member, the *Pax10* Gene

Nathalie Feiner<sup>1,2,3</sup>, Axel Meyer<sup>1,2</sup>, and Shigehiro Kuraku<sup>1,2,4,\*</sup>

<sup>1</sup>Department of Biology, University of Konstanz, Germany

<sup>2</sup>International Max-Planck Research School (IMPRS) for Organismal Biology, University of Konstanz, Germany

<sup>3</sup>Present address: Department of Zoology, University of Oxford, United Kingdom

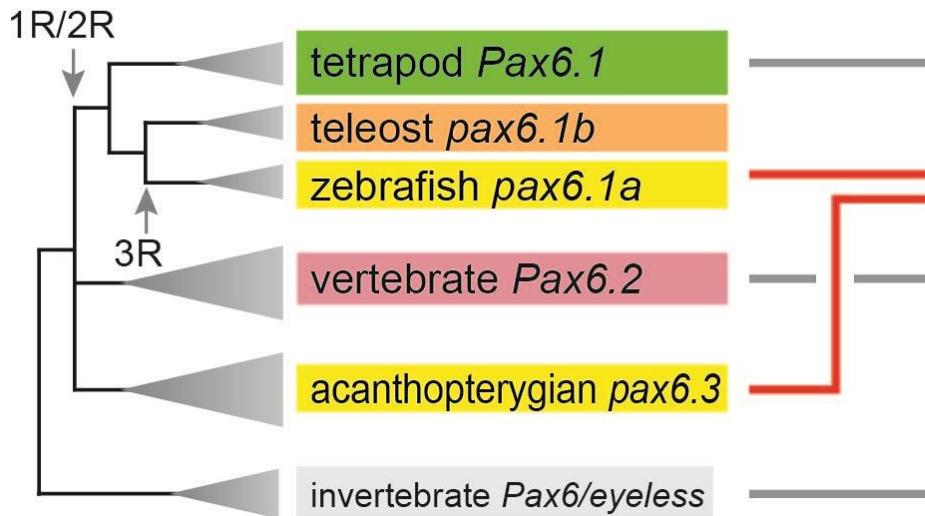
<sup>4</sup>Present address: Genome Resource and Analysis Unit, RIKEN Center for Developmental Biology, Chuo-ku, Kobe, Hyogo, Japan

\*Corresponding author: E-mail: shigehiro-kuraku@cdb.riken.jp.

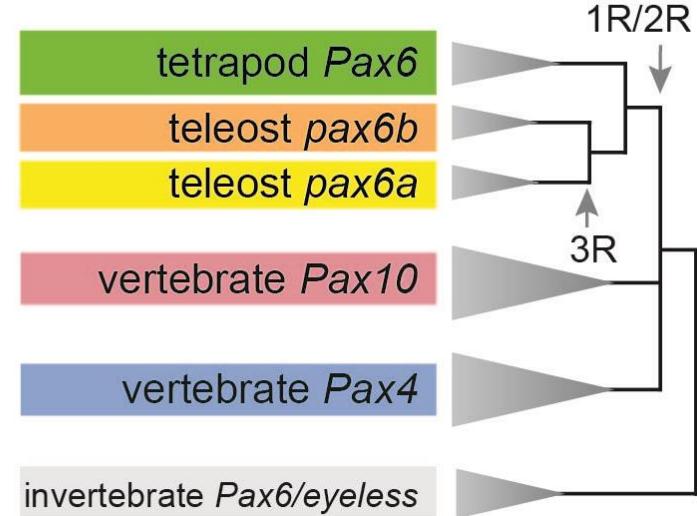
Accepted: June 14, 2014

Data deposition: This project has been deposited at EMBL under the accessions HF567444–HF567455.

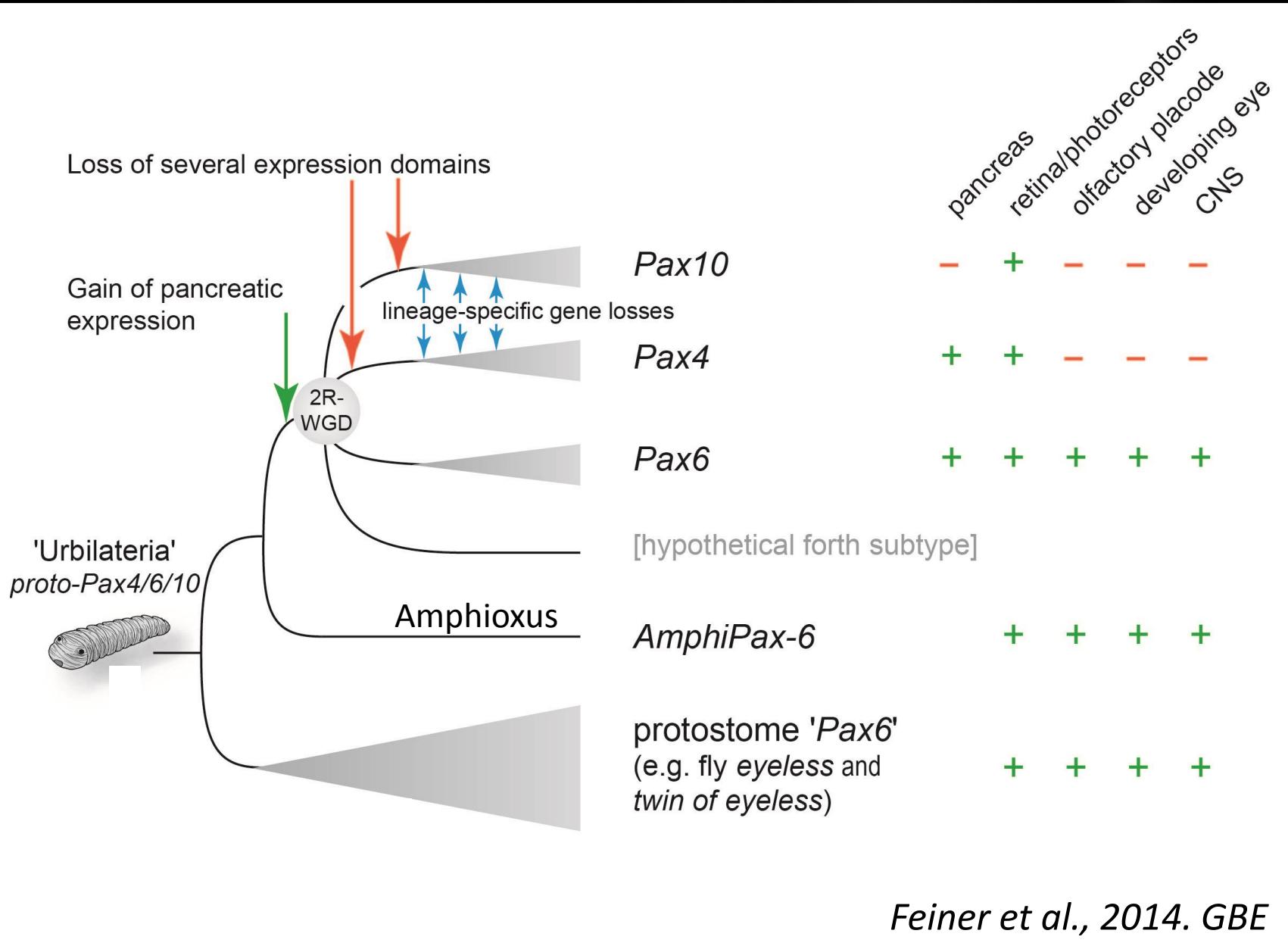
### Hypothesis 1: (Ravi et al., 2013)



### Hypothesis 2: (our study)

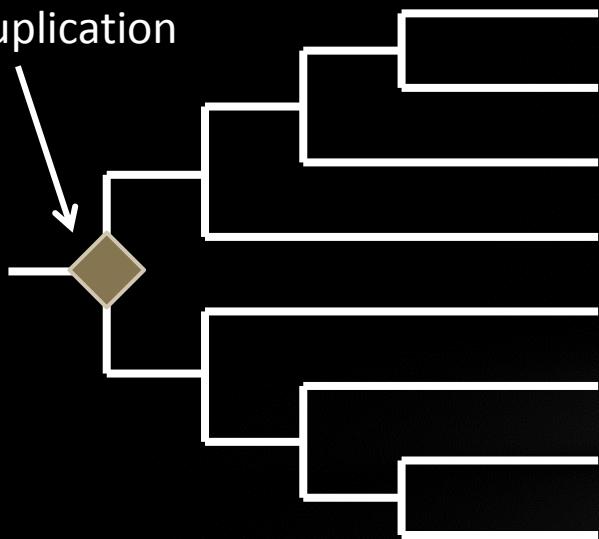


# Revised *Pax6*-*eyeless* relationship

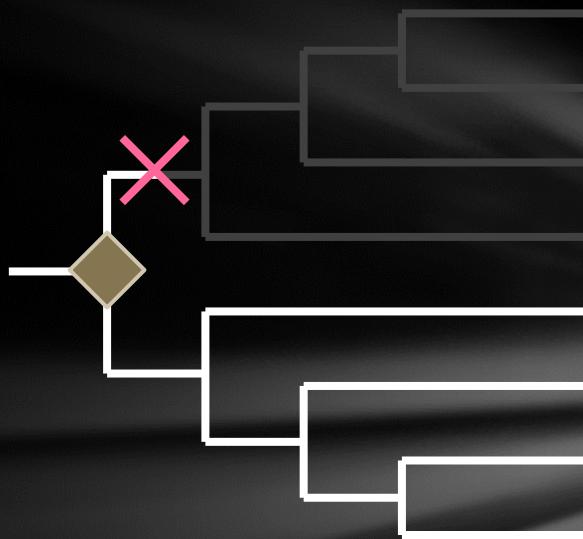


## No loss

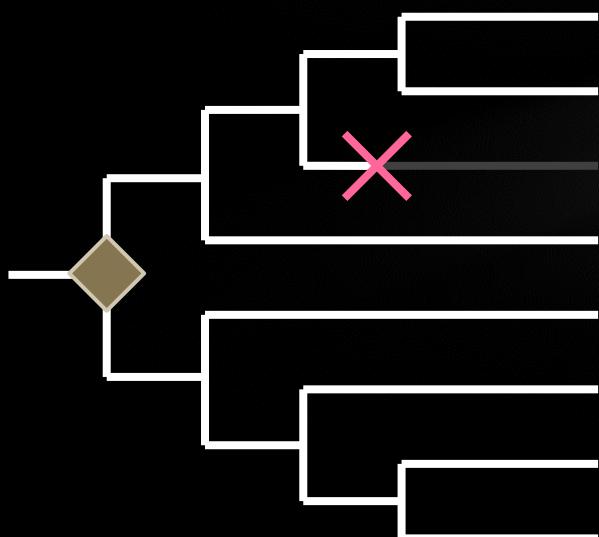
gene duplication



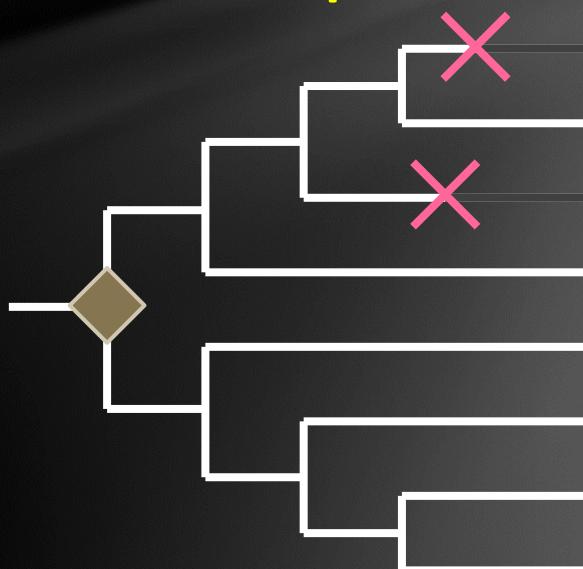
## Loss just after duplication



## Loss long after duplication



## Multiple losses



## No loss

gene duplication

## Loss just after duplication

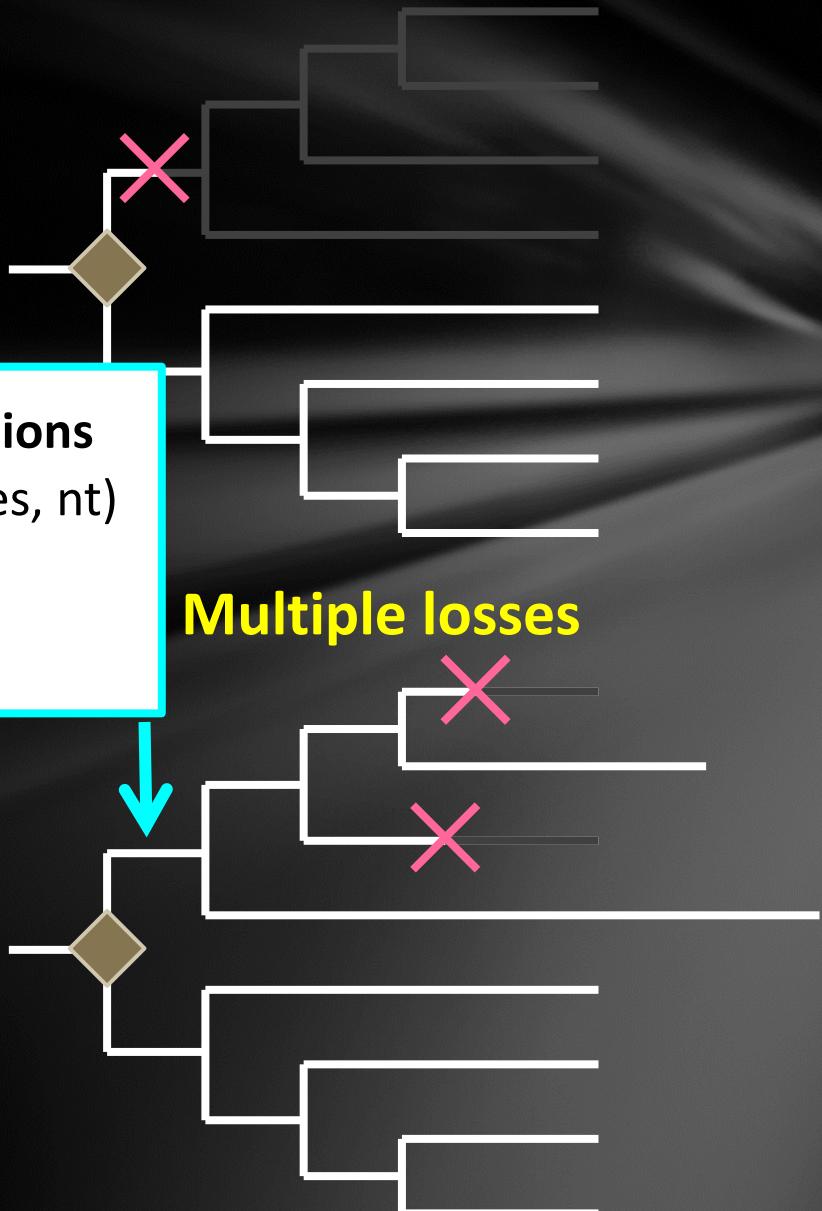
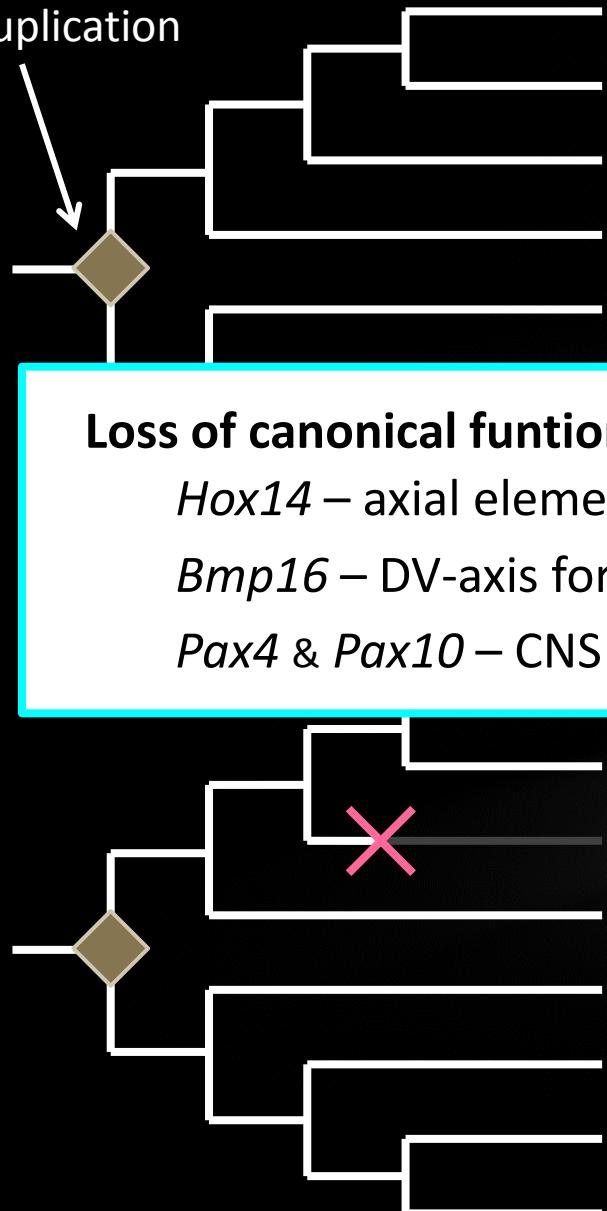
### Loss of canonical funtions/expressions

*Hox14* – axial elements (somites, nt)

*Bmp16* – DV-axis formation

*Pax4* & *Pax10* – CNS

### Multiple losses

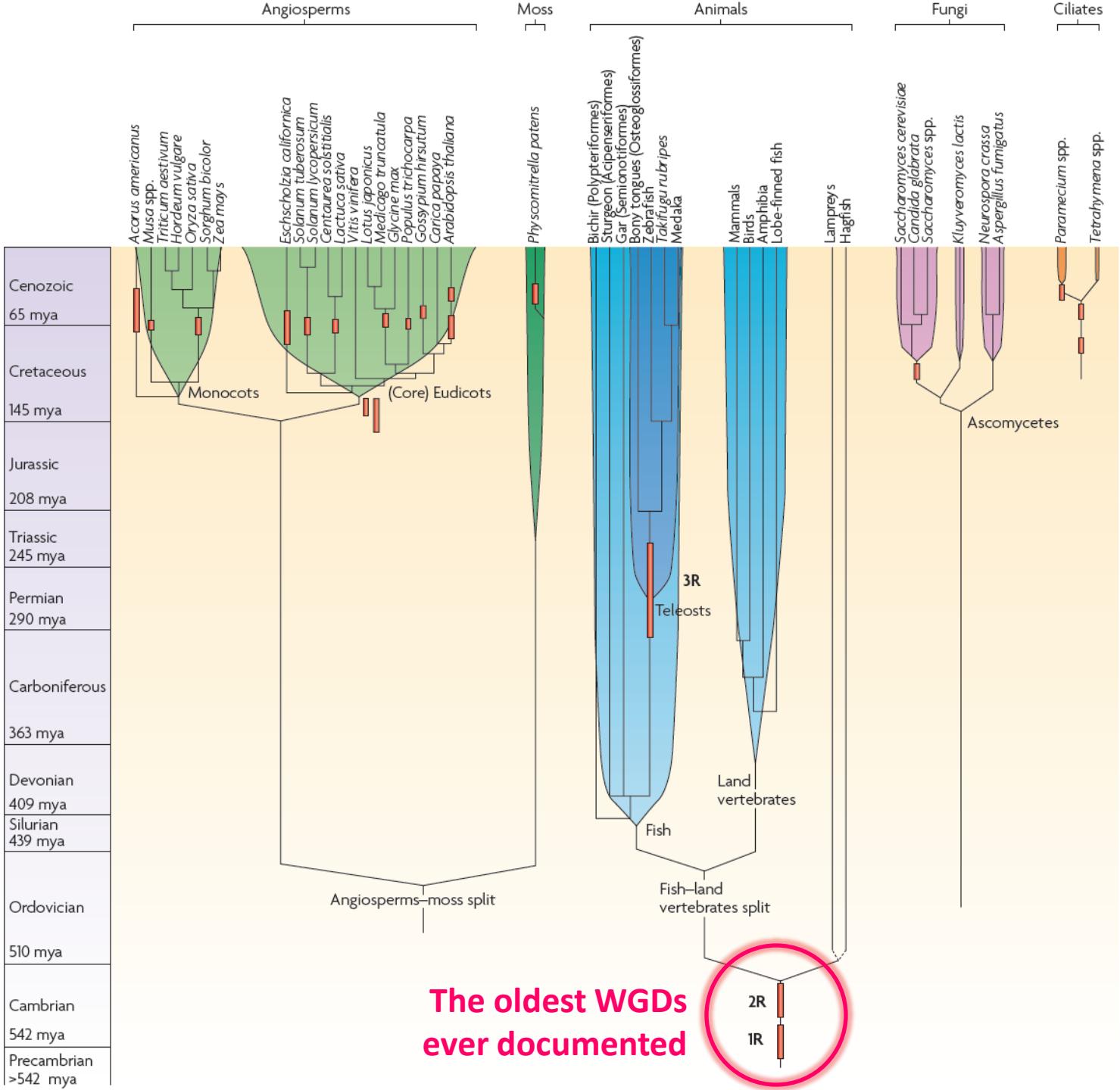


# Agenda

- 1) Revealing non-conservation of developmental ‘toolkits’  
‘Cryptic pan-vertebrate genes’
  
- 2) ‘Two-round’ whole genome duplications  
Warning in using lamprey genomes  
‘lamprey dialect’ misleading phylogeny inference?  
access to less incomplete predicted gene set

# Whole genome duplications (WGDs)

Van de Peer, Maere,  
& Meyer. 2009.  
Nat Rev Genet



# Cyclostomata (hagfishes and lampreys)

Japanese lamprey  
*Lethenteron japonicum*



By K.G. Ota, S. Kuraku and S. Kuratani

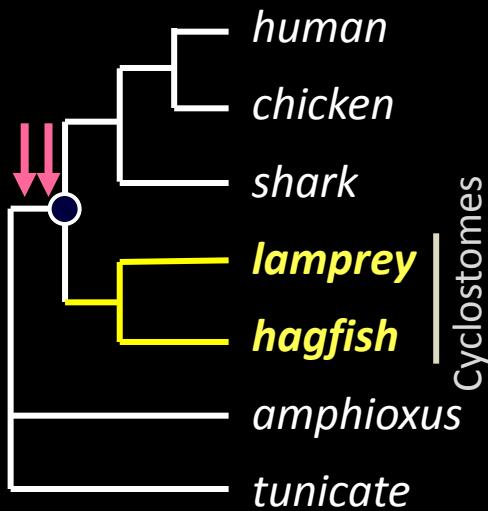
sea lamprey  
*Petromyzon marinus*



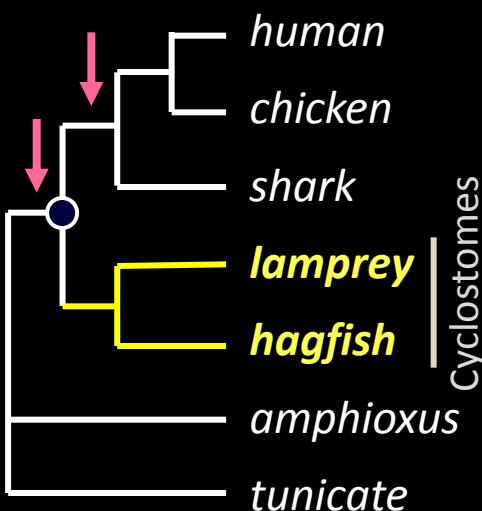
Inshore hagfish  
*Eptatretus burgeri*

# Timing of two-round (2R) WGDs

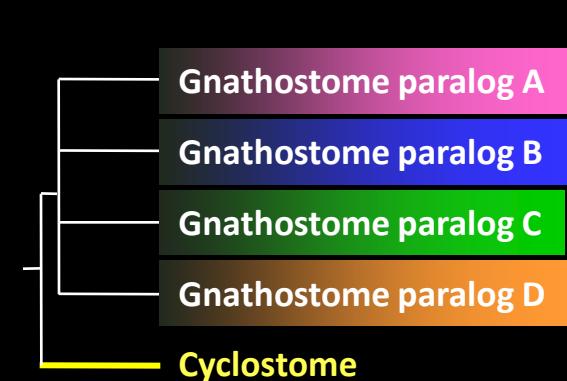
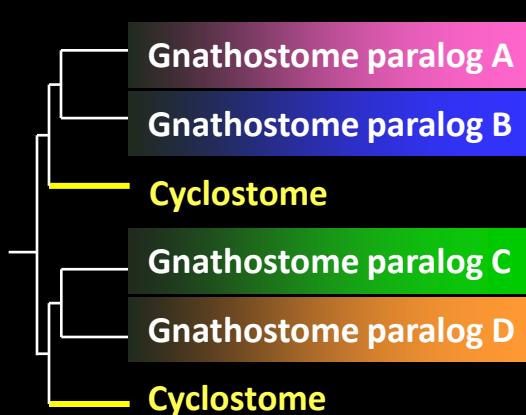
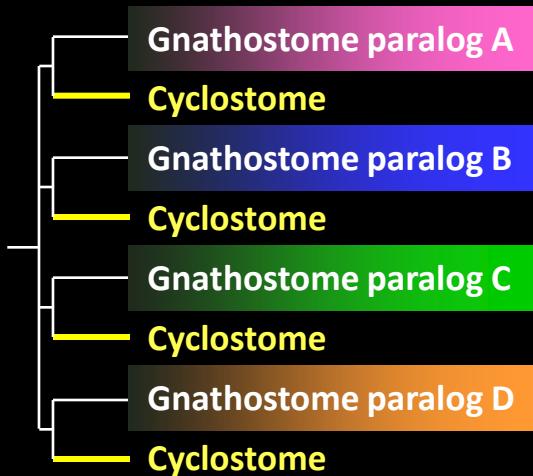
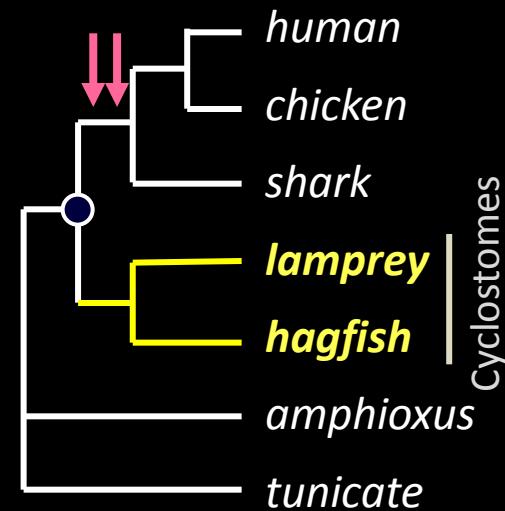
Hypothesis A



Hypothesis B



Hypothesis C



# Timing of two-round (2R) WGDs

Firm evidence of ‘2R’ from tetra-synteny blocks

*Larhammar et al., 2002. Genome Res., etc.*

Independent analyses under different methodologies

2009 Integrative analysis (55 gene families)

*Kuraku et al., 2009. Mol. Biol. Evol.*

2013 Two lamprey genomes

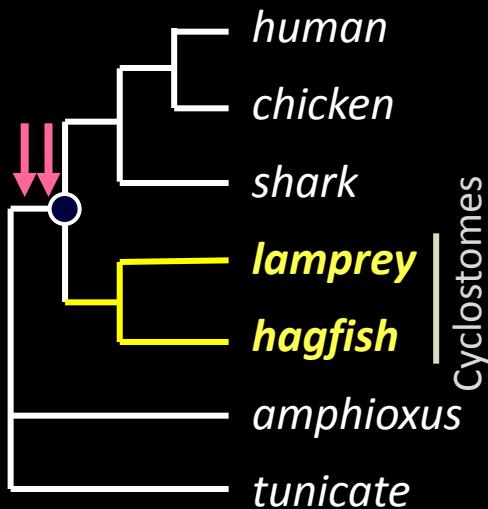
*Petromyzon marinus* (Smith, Kuraku, et al., 2013. Nature Genet)

*Lethenteron japonicum* (Mehta et al., 2013. PNAS)

? Phylome analysis with dense taxon-sampling ?

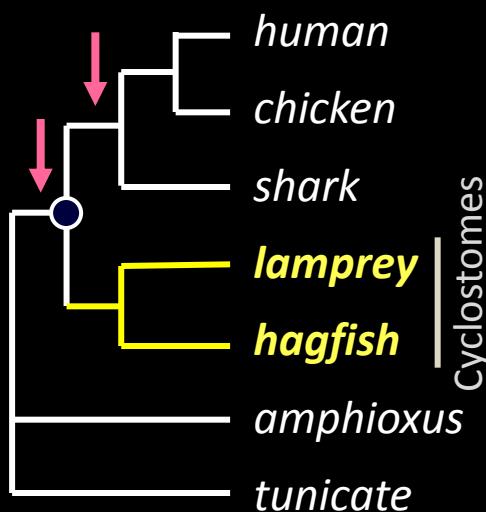
# Timing of two-round (2R) WGDs

## Hypothesis A



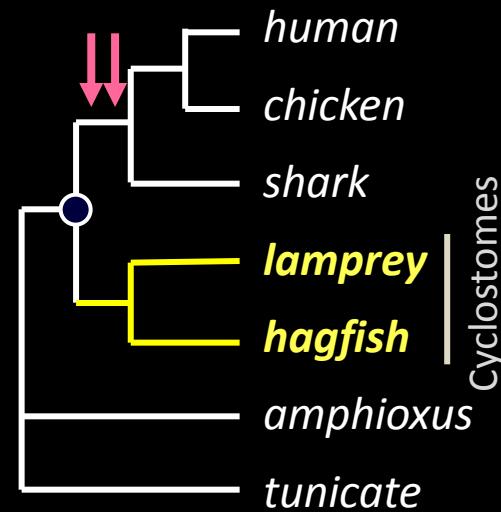
- Mol. phylogeny of 55 gene families  
*Kuraku et al., 2009. MBE*
- Globin gene phylogeny  
*Hoffmann et al., 2010. PNAS*
- Sea lamprey genome analysis  
*Smith, Kuraku et al., 2013. Nature Genetics*

## Hypothesis B



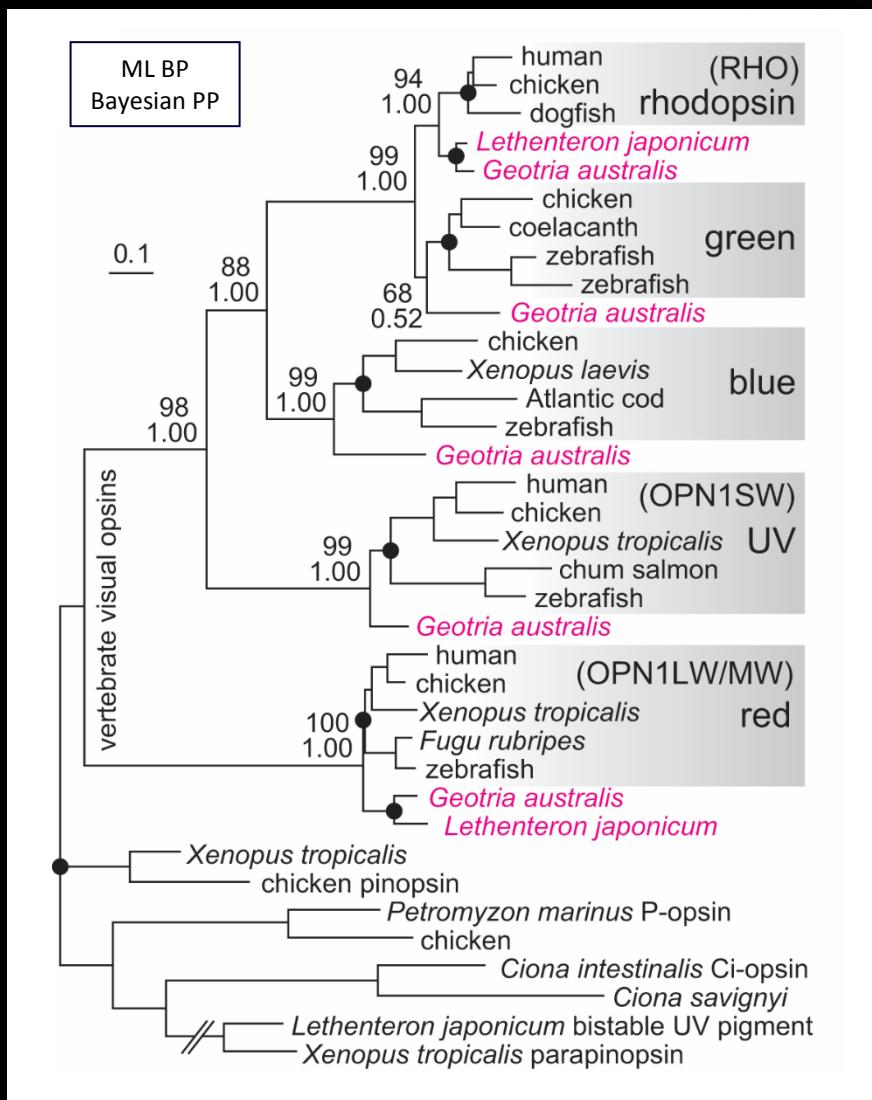
- Composition of Hox/Dlx clusters  
*Neidert et al., 2001. PNAS*  
*Irvine et al., 2002. J Exp Zool B*  
*Force et al., 2002. J Exp Zool B etc*
- Mol. phylogeny of 33 gene families  
*Escriva et al., 2002. MBE*
- Amphioxus genome  
*Putnam et al., 2008. Nature*

## Hypothesis C



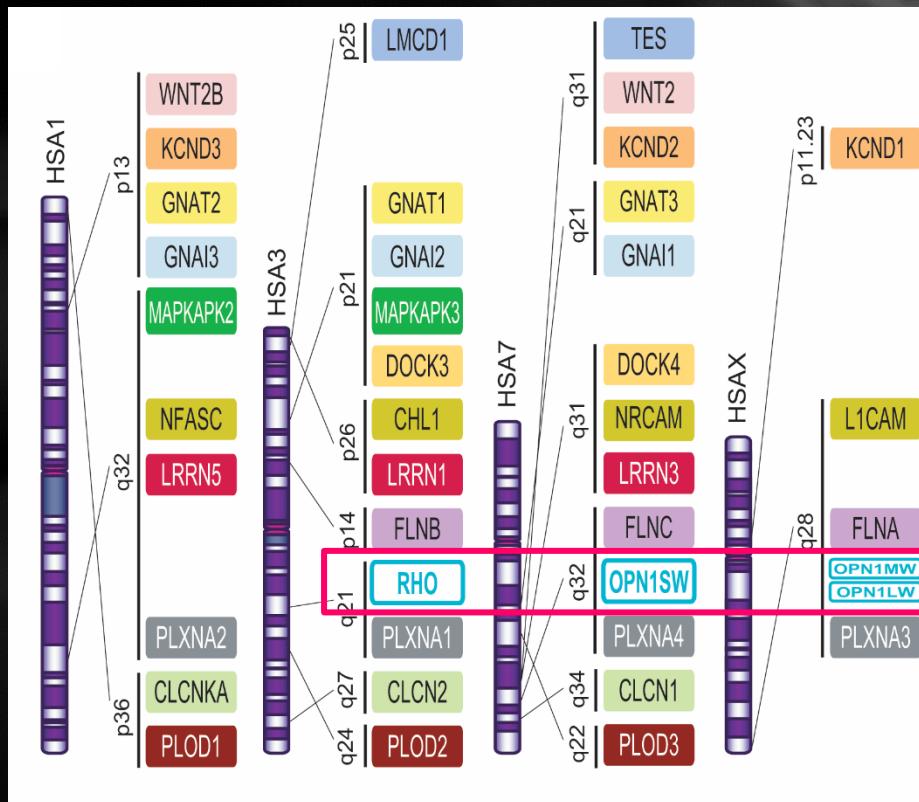
- ParaHox clusters  
*Furlong et al., 2007. MBE*

# Visual opsins



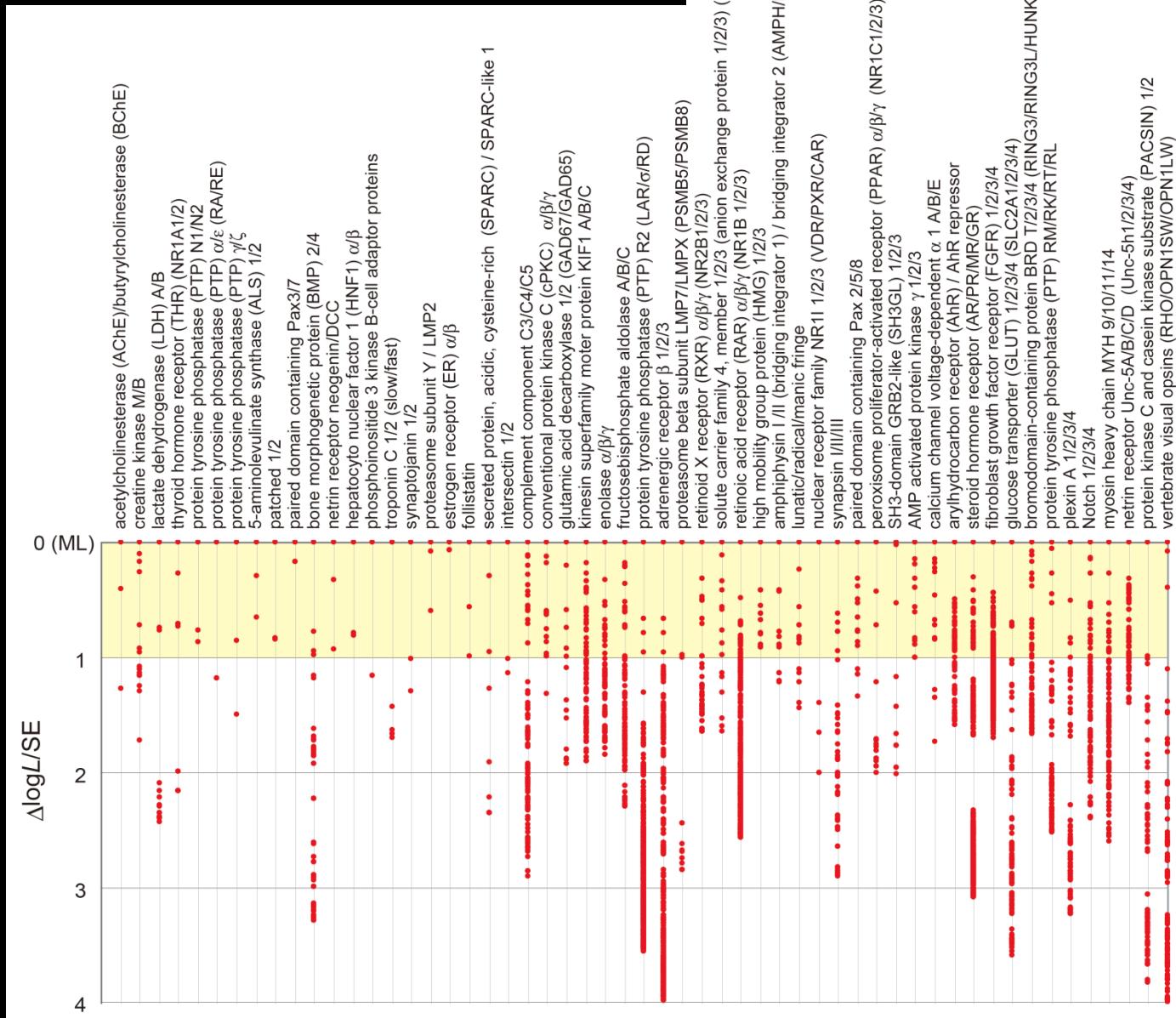
Exhaustive ML, 11 OTUs  
 $JTT+\Gamma_4$  ( $\alpha=0.92$ ), 280 sites

Kuraku et al., 2009. *Mol. Biol. Evol.* 26:47-



1:1 relationships between cyclostome and gnathostome genes

# Not only one tree topology supported

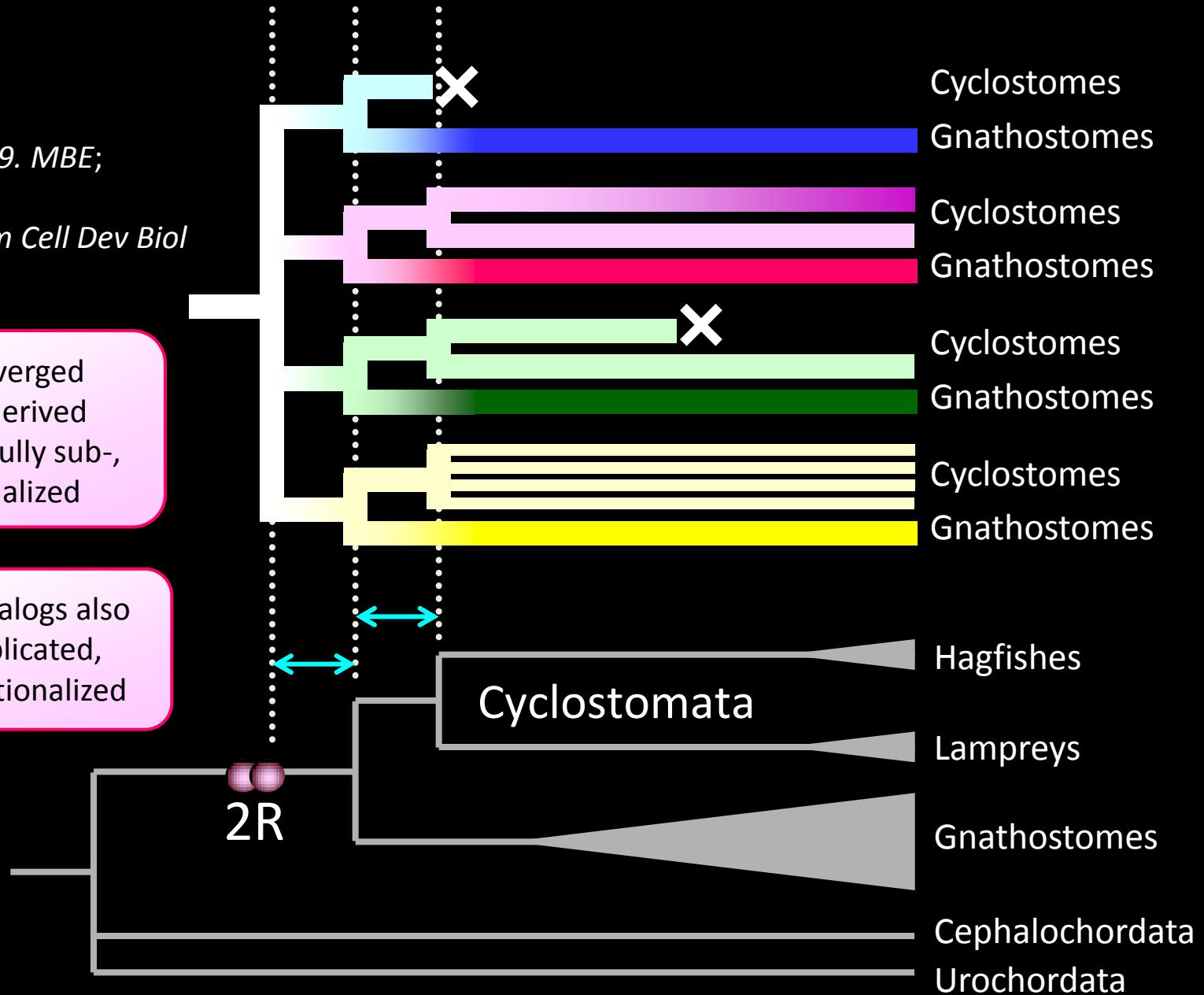


# What Characterizes 'Cyclostomata' ?

Kuraku et al., 2009. MBE;  
Reviewed in  
Kuraku, 2013. *Sem Cell Dev Biol*

Cyclostomes diverged before the 2R-derived paralogs were fully sub-, or neo-functionalized

Cyclostome paralogs also secondarily duplicated, lost or neofunctionalized

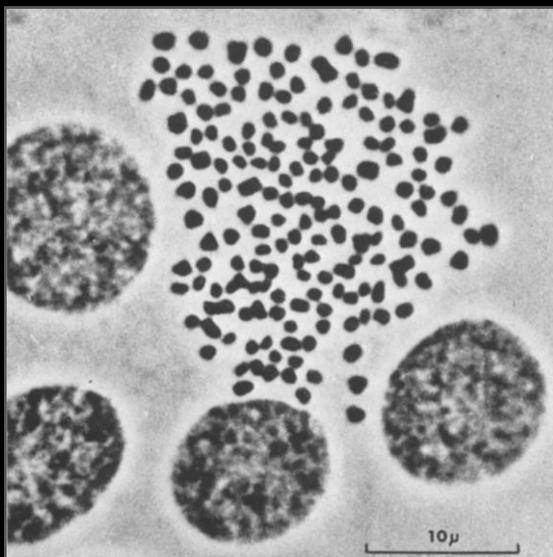




# Sea lamprey genome project

Sequenced at Wash. Univ. Genome Institute

*Petromyzon marinus* (2n=168)



Potter and Rothwell, 1970

## International consortium

Smith, Kuraku, et al. 2013

Nature Genetics

Account · Logout

Search Lamprey...

Lamprey (Pmarinus\_7.0) ▾

**Lamprey**  
*Petromyzon Marinus*

Search Lamprey...  Go

e.g. O42160\_PETMA or GL476598:172728-528162 or prion

Genome assembly: Pmarinus\_7.0

More information and statistics

Download DNA sequence (FASTA)

Example region

Comparative genomics

What can I find? Homologues, gene trees, and whole genome alignments across multiple species.

More about comparative analysis

Download alignments (EMF)

Example gene tree

Gene annotation

What can I find? Protein-coding and non-coding genes, splice variants, cDNA and protein sequences, non-coding RNAs.

More about this genebuild

Download genes, cDNAs, ncRNA, proteins (FASTA)

Update your old Ensembl IDs

Pax6 INS FGF8 BRCA2 DMD ssh Example gene

Example transcript

Variation

This species currently has no variation database. However you can process your own variants using the Variant Effect Predictor:

Variant Effect Predictor

[http://www.ensembl.org/Petromyzon\\_marinus/Info/Index](http://www.ensembl.org/Petromyzon_marinus/Info/Index)

Coding genes: 10,415

Incomplete genome assembly: Pax6 missing

Incomplete gene annotation: Fgf8/17-A missing

=> Access to the consortium gene set??

# Timing of two-round (2R) WGDs

Firm evidence of ‘2R’ from tetra-synteny blocks

*Larhammar et al., 2002. Genome Res., etc.*

Independent analyses with different methodologies

2009 Integrative analysis (55 gene families)

*Kuraku et al., 2009. Mol. Biol. Evol.*

2013 Two lamprey genomes

*Petromyzon marinus* (Smith, Kuraku, et al., 2013. Nature Genet)

*Lethenteron japonicum* (Mehta et al., 2013. PNAS)

Analyzing systematic bias, etc.

? Phylome analysis with dense taxon-sampling ?



# Lamprey genes/genomes

## Protein-coding genes

High GC, long UTR, frequent simple repeats ('QQQQ...', 'HHHH...', etc.)

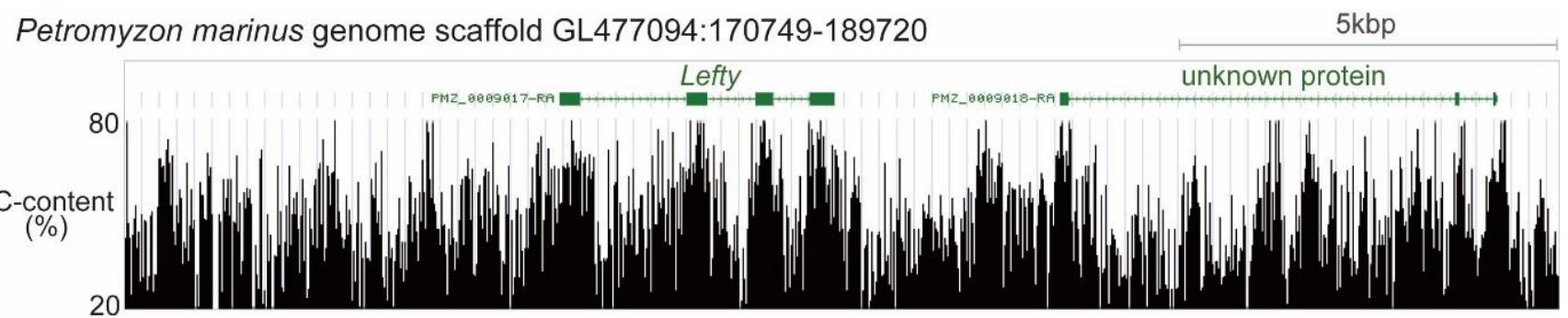
Ambiguous orthology to gnathostome genes

cf. Bmp2/4-B, Dlx-A, Hh-A, HoxW10a, ....

## Genomic landscape

Abundant repetitive elements including foreign Tc1 *Kuraku et al., 2012*

High GC in exons

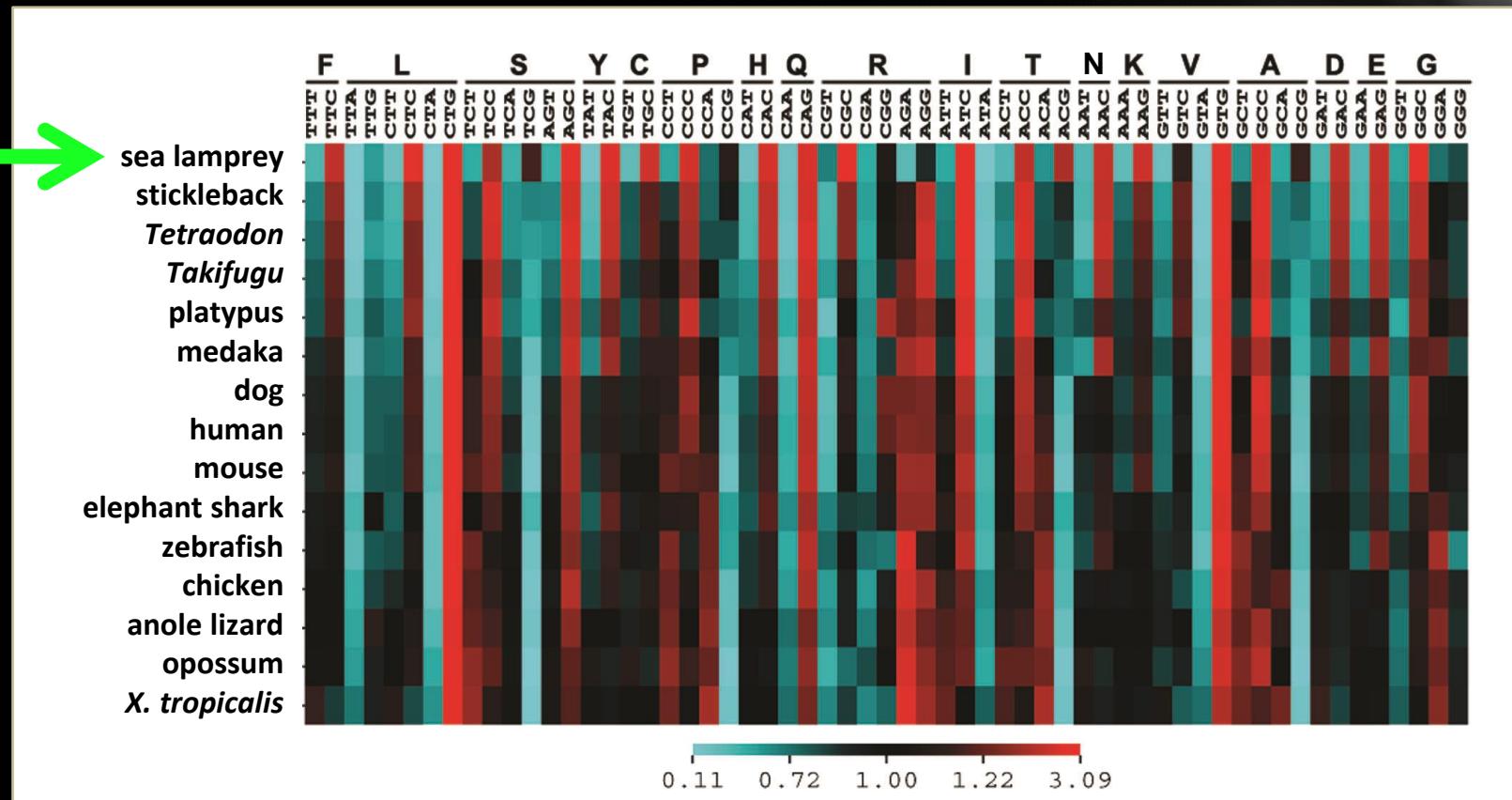




# Peculiar nature of lamprey genes

## Codon usage bias

Methods: RSCU (Sharp et al., 1986) and ENC (Wright, 1990)



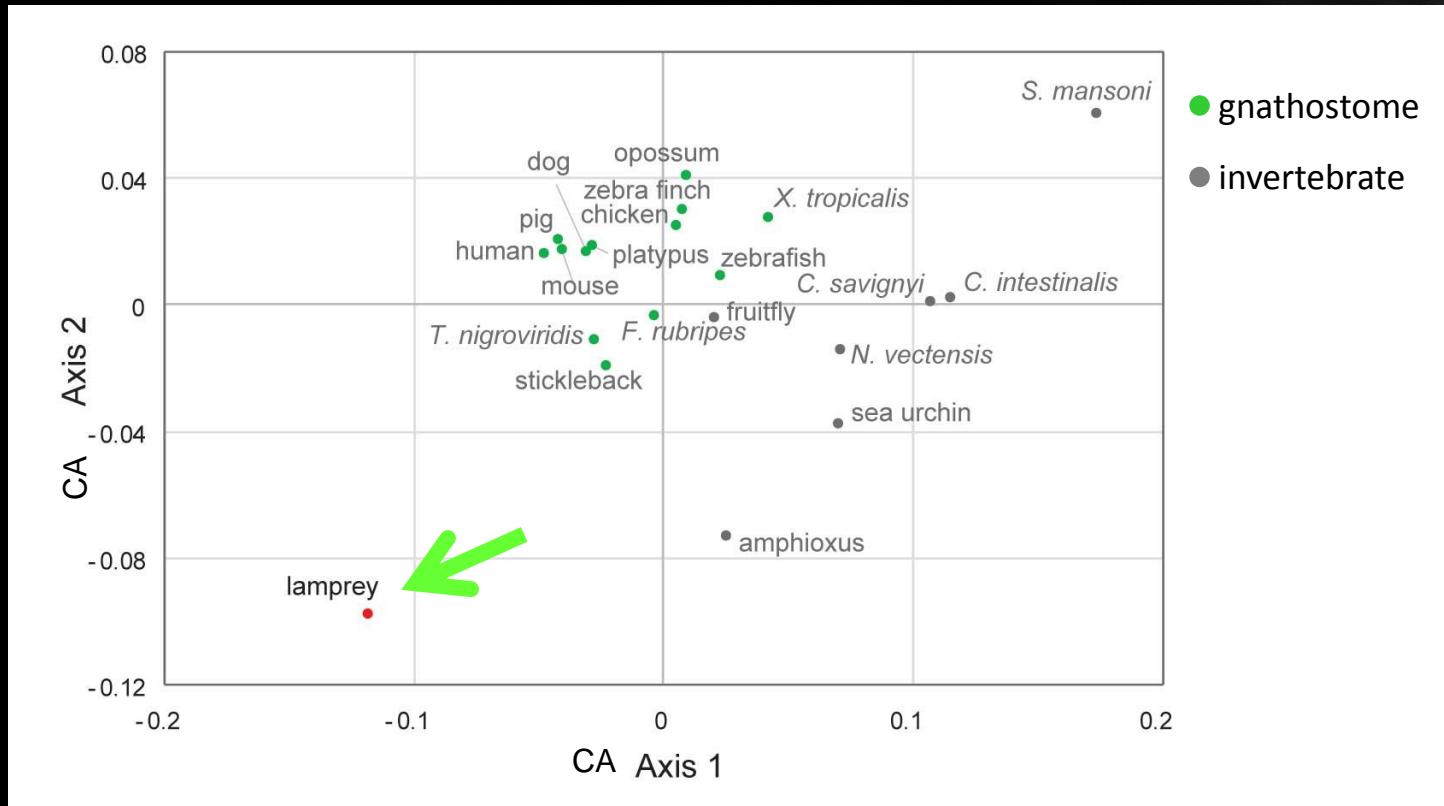
Heavy use of GC-rich codons in translation



# Peculiar nature of lamprey genes

## Amino acid composition

Methods: Correspondence analysis for frequencies of 20 amino acids



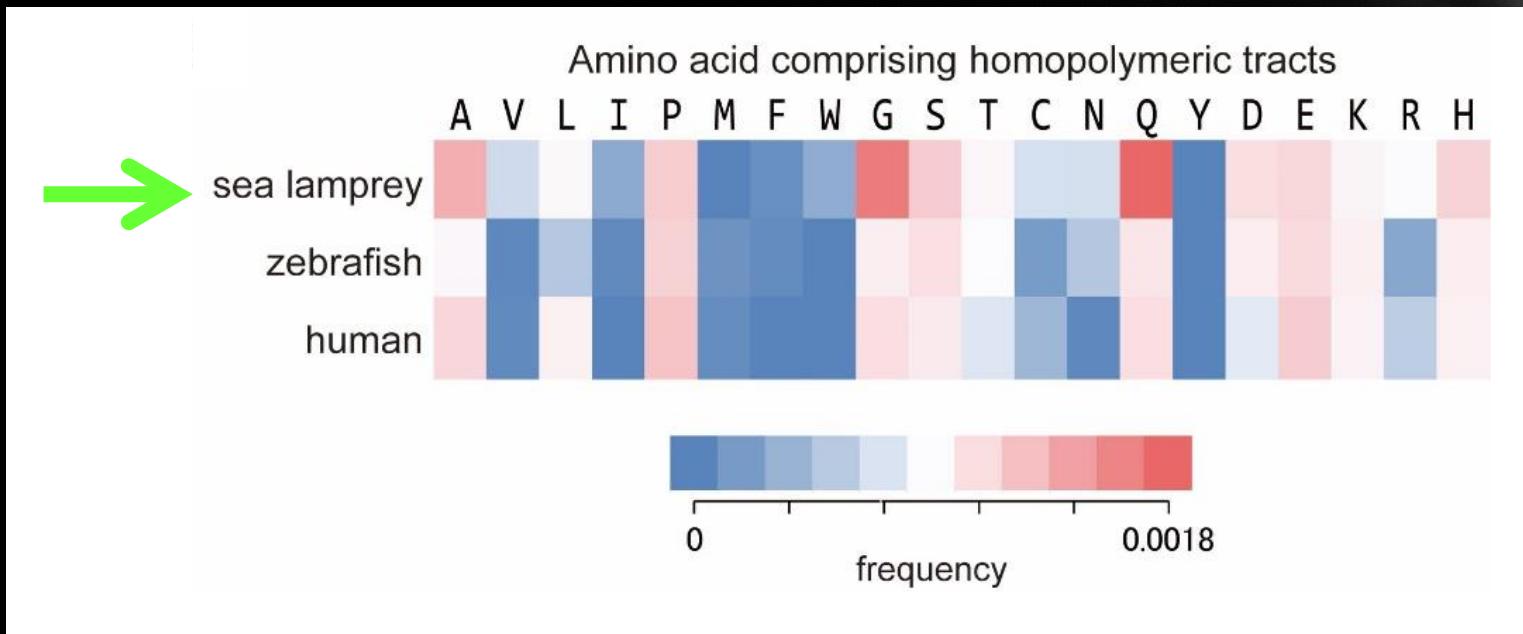
Genome-wide deviation of 'gene model' in lamprey

Smith et al., 2013. Nature Genetics



# Peculiar nature of lamprey genes

Unique pattern of homopolymeric amino acid (HPAA) tracts



# Independent HPAA insertions can mislead phylogeny

cf. Tank EM, ...., Langeland JA. *Evolution & Development* 2009, 11:343-353.

↑  
HPAA  
insertion

*Petromyzon marinus* (sea lamprey) EmxA  
MFQPPTKRCFTIESLVAKDGGDSVAAATL**QQQQQQQQQQQQQQQQQQ**PAYPIPGGANPL  
TCGAQPPHPF**AAAAAAAAAAAAA**SRAGMPELFHDAHLHQPLLA  
PPSLPLFGGPQGRDAISLYPWLHRPRYLGHRYPGADGNAESLLLHSPFARK**PKRIRTAF**  
**SPSQLLRLEHAFEKHNHYVVGAERKQLASSLSLSETQVKWFQNRRTKYKRQKLEE**EGPESP  
QKKKSSHINRWRQATNQGSGDEIDVTSDD ← homeodomain

↑  
HPAA  
insertion

*Petromyzon marinus* (sea lamprey) EmxB  
MFQPATTKRCFTIESLVAKDCPAPTSRSE**QQQQQQQQQQ**HEAPLRPAALSFAGTHNHQHHQ  
HQQHHHHSSASVS**AAAAAAAAAAAAA**FVPAFPQPTARALYPHPAELLSDPGHHPGSAGG  
PLQVPALPPHHHLQSHPLFGPPQRDPMTFYPWLLNRHRYLGHRYPGPETGHEGLLFPGPL  
ARK**PKRIRTAFSPSQLRLLEHAFEKHNHYVVGSERKQLASSLSLSETQVKWFQNRRTKHKR**  
**QKLEEEGPDEQQKKGTTHVNRWRMATKQPSSEDIDVTSDD**

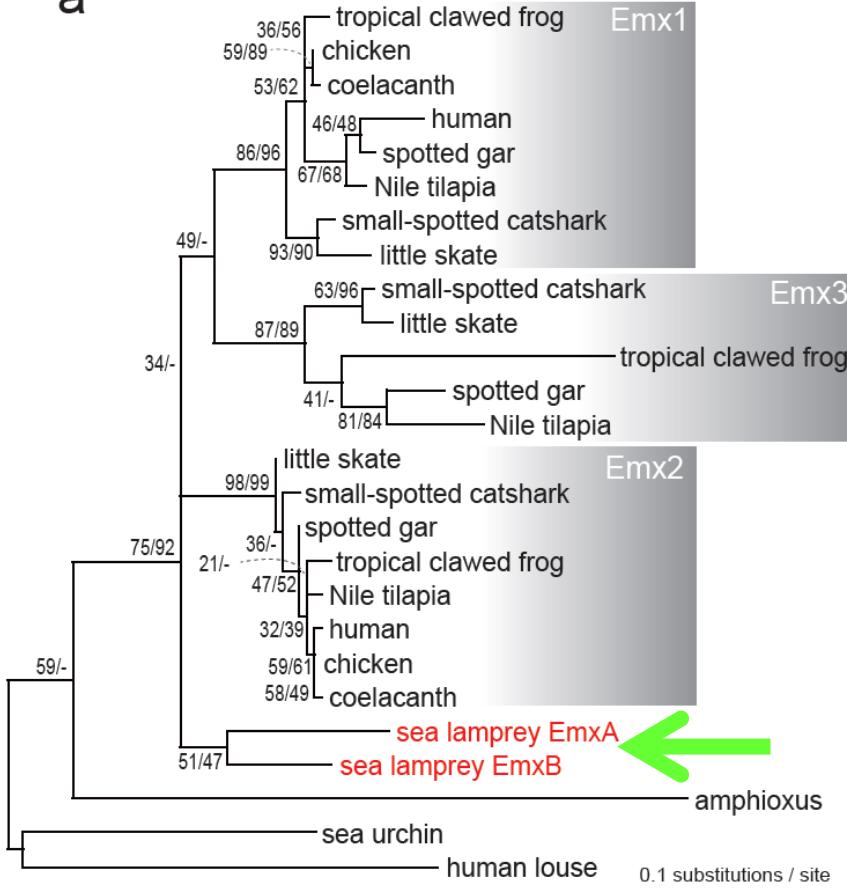
*Eptatretus burgeri* (inshore hagfish) EmxB  
MFQPAAKRCFTIESLVAKDCPAPGGASNRPDEPLRPAALSFAGAPVHSQPGTAFMPGFAQP  
AGRTLYPHADLMYPEPGPHAAASGPLPIPALPVGAHPLQPSHHPLFGPPQRDPMTFYPWLL  
NRHRYLSHRYPGPDSCAESLLFPGPFARK**PKRIRTAFSPSQLRLLEHAFEKHNHYVVGSERK**  
**QKLEEEGPDEQQKKGTTHVNRWRMATKQPSSEDIDVTSDD**

# EmxA-B proximity supported only for lamprey

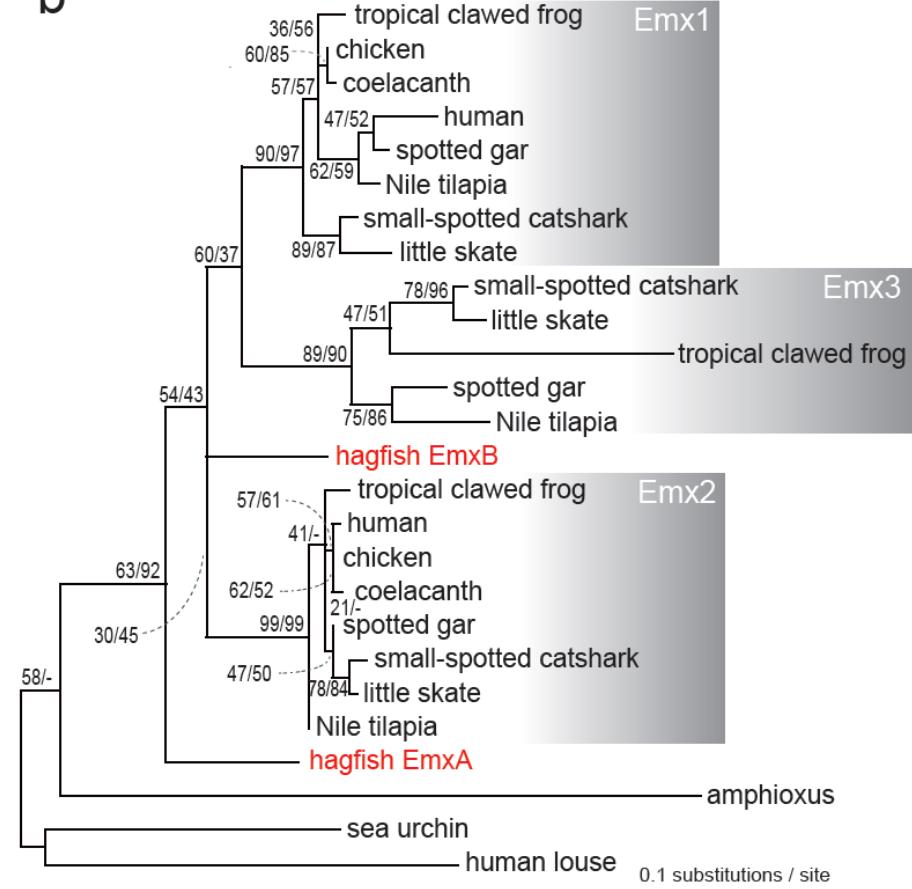
Tree inference with hagfish and/or lamprey

Various alignment and tree inference programs tested

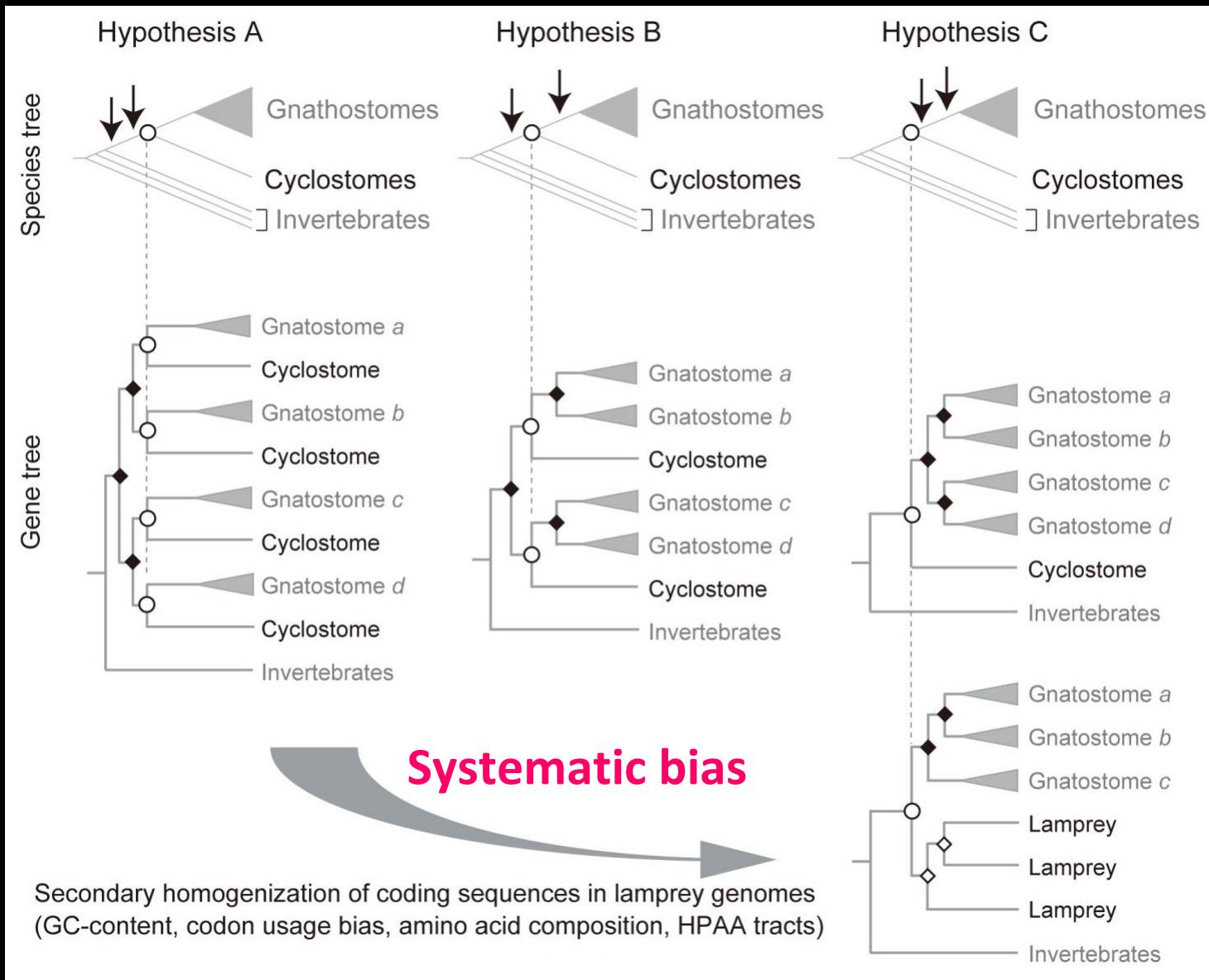
a



b



# Why highly incongruent ?



## **Arising questions and concerns**

Why wasn't QfO initiated earlier?

Any clever way to cope with 'lamprey dialect'?

'Non-homogeneous' methods?

More taxon sampling?

# Taxon sampling



Species in yellow - Currently sequenced in-house

## Arising questions and concerns

Why wasn't QfO initiated earlier?

Any clever way to cope with 'lamprey dialect'?

'Non-homogeneous' methods?

More taxon sampling?

Basically all cyclostome (lamprey and hagfish) genes have ambiguity in orthology

This is also true for genes of eel, arowana, etc. in relation to '3R' (teleost fish-specific) WGD

reviewed in *Kuraku, 2013. Sem Cell Dev Biol*

How do we support and monitor 'quest by non-experts'?

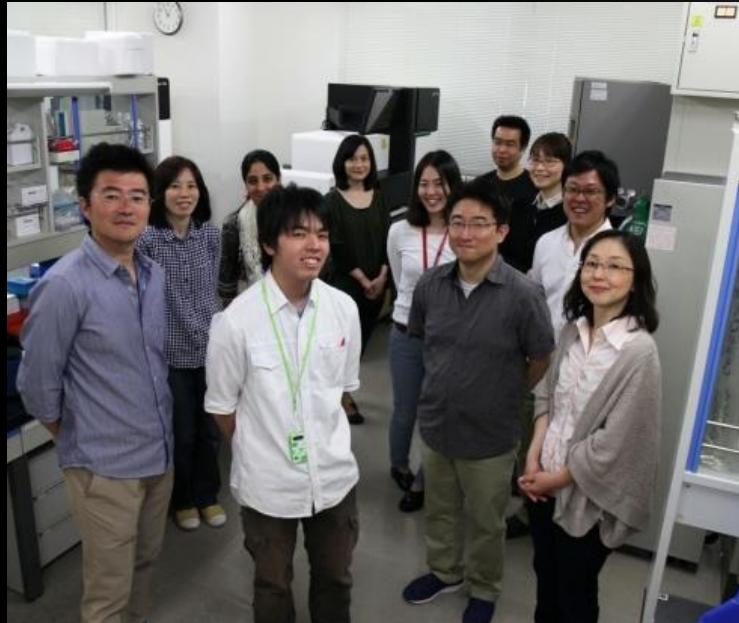
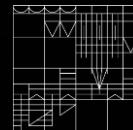
# Acknowledgements

**Sea Lamprey Genome Consortium**

**Meyer Lab in Univ. Konstanz**

**Kuratani Lab in RIKEN Kobe**

**Lab Members in RIKEN Kobe (since March '12)**



Osamu Nishimura  
Chiharu Tanegashima  
Kaori Tatsumi  
Munazah Andrabi  
Yuichiro Hara  
Mitsutaka Kadota  
Kaori Tanaka  
Sayo Saito  
Fumio Motone  
Yuka Kageyama

***Alumni***  
Kazu Itomi  
Tetsutaro Hayashi  
Miyuki Noro  
Kana Miyagishi  
Kenichiro Uno

Open to consultation about NGS  
Possibility for Ph.D. and postdoc